Design and characterization of miconazole nitrate loaded nanosponges containing vaginal gels

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ABSTRACT

The objective of the present work is to prepare and evaluate vaginal gels incorporating nanosponges of Miconazole nitrate for systemic delivery of the drug after topical application. Hence efforts were made to prepare miconazole nitrate loaded nanosponges containing vaginal gels using polymers like hydroxy propyl methyl cellulose (HPMC), sodium carboxy methyl cellulose (NaCMC), methyl cellulose (MC) and carbopol. The gel formulations were prepared with a view to improve permeability of drug. The prepared gel were evaluated for pH, Viscosity, Spreadability, Extrudability, Mucoadhesive time and Invitro diffusion study. The gel formulations can be graded in the following order with respect to the rates of diffusion of drug from them: (HPMC) > (Carbopol) > (NaCMC) > (MC). The correlation coefficient values (r) revealed that the diffusion profiles follows zero order kinetics and the mechanism of drug release was governed by peppas model. The diffusion exponent of release profiles (slope) has a value of (n>0.5), which indicates non fickian diffusion. It was found that the miconazole nitrate loaded nanosponges containing gels prepared with hydroxy propyl methyl cellulose showed good extrudability, homogeneity, spreadability and required diffusion rate in comparison with other formulations and was selected as suitable candidate to be delivered through vaginal route at controlled rate.

INTRODUCTION

Miconazole nitrate is a broad-spectrum antifungal agent of the imidazole group. It acts by means of a combination of two mechanisms: ergosterol biosynthesis inhibition, which causes lysis of fungal cell membranes because of the changes in both membrane integrity and fluidity and direct membrane damage of the fungal cells. The drug is used as a topical treatment for cutaneous mycoses. Poor dissolution and lack of absorption make it a poor candidate for oral administration. Miconazole nitrate poor skin-penetration capability presents a problem in the treatment of cutaneous diseases by topical application. For effective treatment, various approaches have been used to enhance the access of
such poorly skin-partitioned drug molecules. One of the interesting features is the possibility of vaginal use, for which the systems have to be incorporated into commonly used dermal carriers, such as gels, in order to have a proper semisolid consistency [1].

These vaginal formulations are associated with limitations such as poor retention, leakage and messiness, thereby causing inconvenience for users. To overcome these limitations, formulations that adhere to vaginal mucosa for a sufficient period of time need to be developed. Bioadhesion and prolonged retention are desirable characteristics that can be built in vaginal formulation by the use of bioadhesive polymers. Hence, the vaginal route of administration offers a promising option for local and systemic delivery of drugs with the use of bioadhesive polymers [2]. Vaginal gels has advantages wide acceptability, feasibility and low cost. Mucoadhesive polymers of natural, semisynthetic or synthetic origin are able to form hydrogels. In the simplest case the drug is dispersed in a mucoadhesive polymer which swells in the presence of biological fluid and exhibits bioadhesive properties. [3] Vaginal gels are known to possess a higher biocompatibility and bioadhesivity and can be rapidly eliminated through normal catabolic pathways, decreasing the risk for irritative or allergic host reaction at the application site. In the present study Miconazole nitrate loaded nano sponges containing vaginal gels in the form of different gels using various bioadhesive polymers Carbopol 934, cellulose derivative hydroxy propyl methyl cellulose (HPMC), sodium carboxy methyl cellulose (NaCMC) and methyl cellulose (MC) were prepared and evaluated.

**MATERIALS AND METHODS**

Miconazole nitrate was obtained as a gift sample from NatcoPharma, Hyderabad. Sodium CMC (200-300cPs, S. D. fine-chem Ltd.; Mumbai), Carbopol 934(Arihanth traders; Mumbai) HydroxyPropylMethyl cellulose (50cPs S. D. fine-chem Ltd.; Mumbai) and Methyl cellulose (28-32%, S. D. fine-chem Ltd.; Mumbai) obtained commercially. All other materials used were of analytical grade.

**Synthesis of β-cyclodextrin nanosponges**

β-cyclodextrin based nanosponges was prepared using Di phenyl carbonate as a cross-linker. Nanosponges were prepared by using β-cyclodextrin and Di phenyl carbonate in 1:1 ratio. Finely homogenized anhydrous β-cyclodextrin and Di phenyl carbonate were placed in a 100 ml conical flask. The system was gradually heated to 100 °C under magnetic stirring, and left to react for 5 h. During the reaction crystals of phenol appeared at the neck of the flask. The reaction mixture was left to cool and product obtained was broken up roughly. The solid was repeatedly washed with distilled water to remove unreacted β-cyclodextrin and then with acetone, to remove the unreacted Di phenyl carbonate and the phenol present as by-product of the reaction. After purification, nanosponges were stored at 25 °C until further use [4].

**Preparation of Miconazole nitrate loaded nanosponges**

The Miconazole nitrate loading into β-cyclodextrin nanosponges was carried out by solvent evaporation technique. In chloroform required quantity of Miconazole nitrate was dissolved separately to form solutions. To the each solution, required quantity of nanosponges will be added and triturated until the solvent evaporated. While triturating the clumps of nanosponges will be segregated and absorbs the drug solubilised solvent. The solid dispersions were dried in an oven overnight (at 50 °C at atmospheric pressure) to remove any traces of solvents and were sieved through 60 # and used for further work [5].

**Preparation of Miconazole nitrate nanosuspension**

The dried drug encapsulated nanosponges were collected and required quantities of drug equivalent nanosponges were transferred into 250ml volumetric flask containing 100ml methanol in order to remove the free unencapsulated drug by solubilising in the methanol. The drug encapsulated nanosponges were separated from the free drug by membrane filtration by using 0.22µ membrane filter. The residual drug loaded nanosponges were collected and dispersed in distilled water by using ultra sonication to form a nanosuspension [6].
Formulation of vaginal gels containing Miconazole nitrate loaded nanosponges

Different drug reservoir gels were formulated as per the composition given in Table 2. The required quantities of polymer was weighed and transferred separately into a mortar. It was triturated with 5 ml of water. The previously prepared required drug equivalent nanosuspensions, methylparaben and propylparaben were incorporated into the polymer dispersion slowly with continuous trituration to obtain a gel. In case of carbopol gel, specified amount of carbopol 934 was soaked in 5 ml of water over night. The previously prepared required drug equivalent nanosuspensions, methylparaben and propylparaben were incorporated into the polymer dispersion with stirring at 500 rpm, by a magnetic stirrer for 1 h. The pH of above mixture was adjusted to 4.5 with triethanolamine (0.5%). The gel was transferred in to a measuring cylinder and the volume was made up to 10 ml with distilled water. The resulting gels were filled in collapsible tubes [7].

EVALUATION OF DRUG RESERVOIR GELS

Drug content

The gels (1 gm containing 100 mg of drug) were dissolved in 50 ml of phosphate buffer (pH 4.5) solutions. The absorbance was measured after suitable dilution at 272 nm against the corresponding blank solution. The blank solutions were prepared by with gels free from drug [8].

pH and viscosity

The pH of the dispersion was measured using pH meter (Systronics Digital –DI-707). The pH of the gels were measured before and after the incorporation of the drug. Viscosity of the gels was determined using a Brook field viscometer. In the present study, gels were subjected to a shear rate ranging from 10 and 90%. The rheograms and viscosity were obtained with the software rheocal [9].

Extrudability

Closed collapsible tubes containing gel was pressed firmly at the crimped end. When the cap was removed gel extrudes until pressure is dissipated. The weight in grams that was required for extruding 0.5 cm of ribbon of gel in 10 seconds was determined. The results for all the formulations were recorded as extrusion pressure in grams [10].

Spreadability

Spreadability of formulation was determined with the apparatus proposed & fabricated by Multimeret al. It consists of wooden block provided with two glass slides. Lower slide was fixed on wooden block and upper slide with one end was tied to glass slide and other end tied to weight pan. A gel equivalent to 2.5 g was placed between two slides and 1000 g weight was placed over it for 5 minutes to press the sample to a uniform thickness. 100g of weight was added to pan. The time (in seconds) required to separate the two slides was taken as a measure of spreadability. Shorter time interval to cover the distance of 7.5 cm indicates better spreadability [11].

Preparation of goat vagina

Goat vagina was collected from slaughter house by cervical dislocation. The epidermal skin was carefully removed and rinsed with normal saline to remove any loose materials. The epidermal skin was cut into 4.91 cm² (sq area) length. The epidermal skin was stored in cold (5-8 °C) normal saline solution. [12] The prepared gels were evaluated by studying drug diffusion through goat vagina.

Drug Diffusion Study

Drug diffusion study was conducted using Franz diffusion cell [13]. The receptor compartment was filled with 15 ml of phosphate buffer having pH 4.5 as diffusion media. Goat vagina was mounted on the donor compartment with the help of an adhesive. The gels (1 gm containing 100 mg of Miconazole nitrate) was placed into the donor compartment. Magnetic stirrer was set at 50 rpm and whole assembly was maintained at 32 ± 0.5 °C. The amount of drug released was determined by withdrawing 1 ml of sample at regular time intervals for 12 hours. The volume withdrawn was replaced with equal volume of fresh buffer solution. Samples were analyzed for drug content using a UV spectrophotometer at 272 nm.

The rate and the mechanism of drug diffusion through the prepared gels were analyzed by fitting the diffusion data into [14], zero-order equation, Q=Q₀−k₀t, where Q is the amount of drug diffused at time t, and k₀ is the diffusion rate. First order
equation, $\ln Q = \ln Q_0 - k_1 t$, where $k_1$ is the diffusion rate constant and Higuchi’s equation, $Q = k_2 t^{1/2}$, where $Q$ is the amount of the drug diffusion at time $t$ and $k_2$ is the diffusion rate constant. The diffusion data was further analyzed to define the mechanism of diffusion by applying the diffusion data following the empirical equation, $M_t/M_\infty = K t^n$, where $M_t/M_\infty$ is the fraction of drug diffused at time $t$, $K$ is a constant and $n$ characterizes the mechanism of drug diffusion from the formulations during diffusion process.

RESULTS AND DISCUSSION

β-Cyclodextrin based nanosponges were prepared by cross-linking β- Cyclodextrin with carbonate bonds of di phenyl carbonate in 1:1 ratio. Miconazole nitrate was incorporated into nanosponges by solvent evaporation method by dissolving the drug in chloroform. The formulated nanosponges were incorporated into various gels and evaluated for diffusion studies. In the present study efforts were made to prepare vaginal gels of Miconazole nitrate using polymers like HPMC, NaCMC, M.C and Carbopol. Vaginal gels prepared with carbopol and hydroxy propyl methyl cellulose was found to be white, translucent and homogenous. But gels prepared with sodium carboxy methyl cellulose and methyl cellulose was found to be off white and homogenous.

Drug content values of the formulations were well within the range between 98.72±0.40-99.83±0.14% (Table 2). The pH of all formulations was around the skin pH 4.4±0.09 to 4.47±0.04 reflecting no risk of skin irritation which was further confirmed by vaginal irritation testing.

Viscosities of gels were presented in Table 3. All gels were found to exhibit plastic flow. It was observed that the gel formulations showed good extrudability, homogeneity and spreadability and the data was presented in Table 4.

The gels prepared with the methyl cellulose, sodium carboxy methyl cellulose, carbopol and hydroxy propyl methyl cellulose shown drug diffusion for a period of 8.5 hours, 9 hours, 10 hours and 12 hours respectively.

The in vitro diffusion profiles of gels across the goat vagina were showed in Figure1. To ascertain the mechanism of drug diffusion, the diffusion data was analyzed by zero order, first order, and Higuchi and Peppas equations. The correlation coefficient values ($r$) were reported in Table 3. Amount of drug diffused versus time curves exhibited straight line for the formulations and confirmed that the diffusion rate followed zero order release kinetics (Figure.2). Percentage of drug release versus square root of time curves shows linearity and proves that all the formulations followed Peppas model (Figure.3). These values revealed that the diffusion profiles followed zero order kinetics and the mechanism of drug release was governed by peppas model. The diffusion exponent of release profiles (slope) has a values of 0.9112 -1.182 ($n\geq1$), which indicates case II transport diffusion. Among all the formulations the gels prepared with hydroxy propyl methyl cellulose were found to be best formulation.

| Table 1: Composition of Miconazole nitrate Gels Containing Various Polymers |
|--------------------------|-----------------|-----------------|-----------------|-----------------|
| Ingredients               | MNG1            | MNG2            | MNG3            | MNG4            |
| Miconazole nitrate loaded nanosponges (mg) | 3317.11 (100mg) | 3317.11 (100mg) | 3317.11 (100mg) | 3317.11 (100mg) |
| Methyl cellulose (mg)     | 500             |                 |                 |                 |
| Sodium carboxy methyl cellulose (mg) |                 | 500             |                 |                 |
| Carbopol 934 (mg)         |                 |                 | 500             |                 |
| Hydroxy propyl methyl cellulose (mg) | 100             | 100             | 100             | 500             |
| Methylparaben (mg)        |                 | 100             | 100             | 100             |
| Propylparaben (mg)        | 50              | 50              | 50              | 50              |
| Tri ethanolamine (0.5%)   | ---             | ---             | q.s             | ---             |
Table 2: Characteristics of Miconazole nitrate Gels Formulated with Different Polymers

<table>
<thead>
<tr>
<th>Formulation</th>
<th>% Drug content</th>
<th>pH</th>
<th>Viscosity (cps)</th>
<th>Spreadability (gm.cm/sec)</th>
<th>Extrudability (N)</th>
<th>Mucoadhesive time</th>
</tr>
</thead>
<tbody>
<tr>
<td>MNG₁</td>
<td>98.72±0.40</td>
<td>4.42±0.09</td>
<td>3498±31</td>
<td>32.16±1.12</td>
<td>90.14±0.05</td>
<td>&gt; 12</td>
</tr>
<tr>
<td>MNG₂</td>
<td>99.83±0.14</td>
<td>4.47±0.04</td>
<td>3721±17</td>
<td>32.44±1.23</td>
<td>90.28±0.03</td>
<td>&gt; 12</td>
</tr>
<tr>
<td>MNG₃</td>
<td>99.48±0.19</td>
<td>4.46±0.29</td>
<td>3978±36</td>
<td>33.27±1.51</td>
<td>91.64±0.07</td>
<td>&gt; 12</td>
</tr>
<tr>
<td>MNG₄</td>
<td>99.63±0.74</td>
<td>4.43±0.18</td>
<td>4464±28</td>
<td>34.72±1.32</td>
<td>91.76±0.03</td>
<td>&gt; 12</td>
</tr>
</tbody>
</table>

Table 3: In vitro drug release kinetic data of Miconazole nitrate gels containing nanosponges prepared with β-cyclodextrin and Diphenyl carbonate in 1:1 ratios and by using different polymers

<table>
<thead>
<tr>
<th>Formulation Code</th>
<th>Correlation Coefficient Values (r)</th>
<th>Diffusion Rate Constant (mg/hr) Ko</th>
<th>t₅₀%</th>
<th>t₉₀%</th>
<th>n Value</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Zero Order</td>
<td>First Order</td>
<td>Higuchi Model</td>
<td>Peppas Model</td>
<td></td>
</tr>
<tr>
<td>MNG₁</td>
<td>0.9996</td>
<td>0.8056</td>
<td>0.9303</td>
<td>0.9990</td>
<td>11.62</td>
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<tr>
<td>MNG₂</td>
<td>0.9989</td>
<td>0.7971</td>
<td>0.9107</td>
<td>0.9998</td>
<td>10.86</td>
</tr>
<tr>
<td>MNG₃</td>
<td>0.9986</td>
<td>0.8073</td>
<td>0.9214</td>
<td>0.9985</td>
<td>9.80</td>
</tr>
<tr>
<td>MNG₄</td>
<td>0.9983</td>
<td>0.6380</td>
<td>0.9052</td>
<td>0.9991</td>
<td>8.31</td>
</tr>
</tbody>
</table>

Figure 1: Comparative in vitro drug release profiles of Miconazole nitrate gels prepared with different polymers.
(-■-) MG1. Miconazole nitrate gels prepared with Methyl cellulose
(-♦-) MG2. Miconazole nitrate gels prepared with Sodium CarboxyMethyl cellulose
(-▲-) MG3. Miconazole nitrate gels prepared with Carbopol
(-×-) MG4. Miconazole nitrate gels prepared with HPMC

Figure 2: Comparative zero order profiles of Miconazole nitrate gels prepared with different polymers.

(-■-) MG1. Miconazole nitrate gels prepared with Methyl cellulose
(-▲-) MG3. Miconazole nitrate gels prepared with Carbopol
(-♦-) MG2. Miconazole nitrate gels prepared with Sodium CarboxyMethyl cellulose
(-×-) MG4. Miconazole nitrate gels prepared with HPMC
Figure 3: Comparative peppas plots of Miconazole nitrate gels prepared with different polymers.

(-■-) MG1. Miconazole nitrate gels prepared with Methyl cellulose
(-●-) MG2. Miconazole nitrate gels prepared with Sodium CarboxyMethyl cellulose
(-▲-) MG3. Miconazole nitrate gels prepared with Carbopol
(-×-) MG4. Miconazole nitrate gels prepared with HPMC

REFERENCE

