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Research article

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Estimation of olopatadine hydrochloride by RP–HPLC and U.V spectrophotometry method in pure and pharmaceutical formulation

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ABSTRACT

A simple and sensitive Reversed Phase High Performance Liquid Chromatographic method and UV Spectrophotometry has been developed and validated for the of the Olopatadine Hcl in pure and pharmaceutical dosage form. In RP-HPLC Method the separation was carried out using mobile phase consisting of Methanol and Acetonitrile in the ratio of 60:40 (v/v). The column used was inertsil ODS 3V C_{18} , (250 mm x 4.6 mm i.d., 5µm) with flow rate 1.2ml/min using UV detection at 254 nm. The method was linear over a concentration range of $10 - 250\mu g/$. The retention time of 2.857min. Results of analysis were validated statistically and by recovery studies. The mean recovery was 98.2 to101.5. The limit of detection (LOD) and the limit of quantification (LOQ) were found to be 0.024 and 0.075µg/ml. The %RSD for the method precision was found to be less than 2%. To develop simple and economical UV spectrophotometric method for the estimation of Olopatadine in pharmaceutical dosage form available in the market for conjunctivitis. The method was validated as per the ICH guidelines and the results were statistically validated. Linearity was observed in concentration range of $10-60\mu g/ml$ for Olopatadine. The accuracy of the method was evaluated by recovery studies and good recovery results were obtained between 98% to 100% and the relative standard deviation was found to be below 2% . A simple, accurate, sensitive and economical UV-spectrophotometric method for the estimation of Olopatadine pharmaceutical dosage form has been developed which can be employed in the industry for the routine analysis. **Keywords:** Olopatadine (OLO), RP-HPLC, UV spectrophotometry

INTRODUCTION

Olopatadine is chemically $\{(11Z)-11-[3-(dimethyl amino)propylidene]-6,11-dihydro dibenzo [$ *b*,*e* $] oxepin-2-yl<math>\}$ acetic acid Olopatadine hydrochloride is an antihistamine (as well as anticholinergic and mast cell stabilizer), sold as a prescription eye drop (0.2% solution, Pataday (or Patanol S in some countries),

manufactured by Alcon). It is used to treat itching associated with allergic conjunctivitis (eye allergies). Olopatadine hydrochloride 0.1% is sold as Patanol (or Opatanol in some countries). A decongestant nasal spray formulation is sold as Patanase, which was approved by the FDA on April 15, 2008

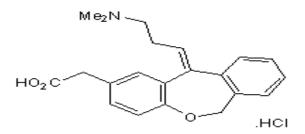


Figure 1: Chemical structure of olopatadine

Several HPLC^{5,6,7}, GC^{8,9} and LC/MS-MS¹⁰⁻¹⁴ methods have been reported for the analysis of olopatadine in plasma that suffer from either undesirably long chromatographic run times and requirement for gradient analysis or use of an internal standard.

The objective of this study was to develop reverse phase high performance liquid chromatography method for the estimation of olopatadine in pure and pharmaceutical dosage form without any derivatization and having short retention time. This method was found to be linear, precise, accurate, sensitive, specific, and robust, and therefore suitable for routine analysis.

MATERIALS AND METHOD

HPLC Instrumentation and Chromatographic conditions

The analytical separations were carried out on a waters 2487 HPLC system equipped with Photo Diode Array detector. The output of signal was monitored and integrated using LC – solutions 2000 software. The analytical column was inertsel ods C_{18} (250 × 4.6mm, 5µ). Mobile phase consisted Acetonitrile and methanol in the ratio of 40:60. Mobile phase was mixed, filtered through 0.45µmembrane filter and degassed under ultrasonication. The mobile phase was used as diluent. The flow rate was 1.0 ml/min and runtime was 5 minute. The column was maintained at ambient temperature. UV detection was measured at 254 nm and the volume of sample injected was 10 µl.

Preparation of standard stock solution

For both uv & rp-hplc method:50mg of olopatadine was weighed accurately and dissolved in 50ml of mobile phase to get the concentration of 1000 μ g/ml. Resultant solution was filtered through Whatman filter

paper. The standard chromatogram for olopatadine $(100\mu g/ml)$ was shown in figure 2.

Preparation of working standard solution For RP-HPLC method

Working standard solutions of olopatadine were prepared by accurately transferring the (0.1, 0.5, 1.0, 1.5, 2.0 and 2.5 ml) aliquots of the standard stock solution into a series of six 10 ml volumetric flasks. The volume was made up to mark with mobile phase to obtain concentration range of 10 - 250 µg/ml.

For UV method

Working standard solutions of olopatadine were prepared by accurately transferring the (0.1, 0.2, .0.3, .0.4, .0.5 and 0.6 ml) aliquots of the standard stock solution into a series of six 10 ml volumetric flasks. The volume was made up to mark with mobile phase to obtain concentration range of $10 - 60 \mu$ g/ml

Preparation of sample solutions For both uv & rp-hplc method

25mg of tablet powder was weighed accurately and was taken into 25mL volumetric flask and then the sample was diluted to 25ml with mobile phase to get concentration of 1000μ g/ml and used for analysis.

RESULTS AND DISCUSSION

HPLC method development and optimization

To optimize the chromatographic conditions, different columns, mobile phases, flow rates etc., were tested. Acetonitrile and water in the ratio of 40:60 was preferred as mobile phase because it resulted in a greater response to olopatadine after several preliminary investigatory runs compared with the different mobile phase combinations. The effect of the flow rate was studied in the range 0.9 to 1.5ml/min and 1.2ml/min was preferred to be effective. Under these conditions, the analyte peak obtained was well-defined

and free from tailing. The retention time (RT) was found to be 2.8min. The optimized chromatographic parameters were listed in table 1

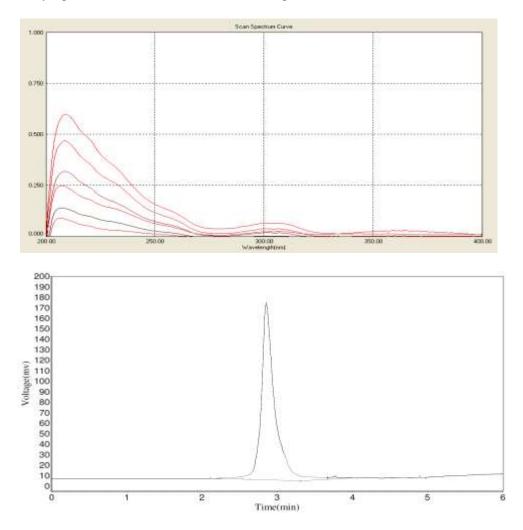


Table 1: Optimized chromatographic parameters

Optimized Chromatographic parameters					
Elution	gradient				
Mobile phase acet	onitrile	:methanol (40:60)			
Column	inertsil c18 c	olumn			
Flow rate	1.2ml/min				
Detection	254nm				
Injection volume	10µl				
Temperature	Ambient				
Retention time	2.87min				
Run time	5.0 min				
Concentration 1	0 - 250µg/ml				

Validation of the method

When method development and optimization are complete, it is necessary to accomplish method validation. The validation studies include linear range (correlation coefficient), method precision (RSD, %), method accuracy (% recovery and RSD, %), sensitivity studies (LOD & LOQ), and robustness.

System suitability studies

System-suitability tests are an integral part of method development and are used to ensure adequate

performance of the chromatographic system. Retention time (RT), tailing factor (T), and peak asymmetry (AS), resolution (RS) were evaluated. The system suitability test was performed using five replicate injections of standards before analysis of samples. The system suitability method acceptance criteria set in each validation run were: capacity factor > 2.0, tailing factor ≤ 2.0 and theoretical plates > 2000. In all cases, the relative standard deviation (R.S.D) for the analytic peak area for two consecutive injections was < 2.0%. System suitability parameters were shown in table 2.

Table 2: System suitability parameters				
Parameters		Values		
Retention time	1.862min			

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Linearity

The linearity of the method was evaluated by preparing six series of standard solutions of olopatadine in the range of $10 - 250 \ \mu\text{g/ml}$ in mobile phase and injecting the solutions into the HPLC system. Excellent

correlation between olopatadine peak area and concentration was observed with $R^2 = 0.998$ (Figure.3). The regression equation was found to be Y =5844 x +75913. Statistical data are presented in table 3 and the calibration curve was shown in figure 3.

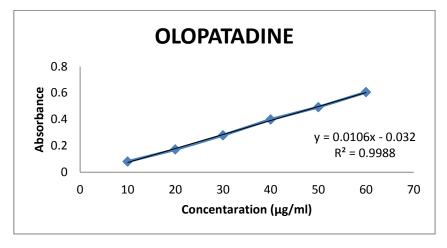
	s.no	concentr	ation	Area		
	1	10	2	417079	_	
	2	50	8	805358		
	3	100	13	3898789		
	4	150	1	974988		
	5	200	2	484887		
	6	250	3	092886		
1200000 - 1000000 - 800000 -	y =	= 5844.8x + R ² = 0.99				
600000 -						
400000 -						
200000 -						
0 +		1	1	1	1	
0	5	0 10	00 15	50	200	250

Figure 3: Calibration curve of olopatadine by RP -HPLC method

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s.no	concentration	Absorbance
1	10	0.081
2	20	0.172
3	30	0.279
4	40	0.399
5	50	0.492
6	60	0.605



Calibration curve for olopatadine by uv method Precision

System precision: (Repeatability)

To study precision, five replicate standard solutions of olopatadine $(100\mu g/ml)$ were prepared and analyzed using the proposed method. The percent relative standard deviation (% RSD) for peak responses was calculated. Results of system precision studies were shown in table 4.

s.no	Retention time(min)	Area(Mv. sec)
1	2.89	1389879
2	2.89	1390870
3	2.94	1381709
4	2.81	1371808
5	2.82	1387968
6	2.857	1375058
mean	2.89	1382882
Standard deviation	0.047545968	8048.07964

Table 4: Results of system precision for olopatadine

% RSD 1.645189498 0.581978769

Method precision: (Reproducibility)

The intraday and inter-day precision of the proposed method was determined by analyzing the corresponding responses 6 times on the same day and on different days for concentration of sample solutions of 100μ g/ml. The result was reported in terms of relative standard deviation (% RSD). Results of method precision studies were shown in table 5.

	1	1 0
s.no	Peak area	%labelled claim
1	1375623	100.5276
2	1367299	101.139
3	1381142	100.125
4	1358812	101.771
5	1385642	99.8008
6	1399221	98.8322

25µg/ml	test abs	std abs	wt to be taken	wt taken	avg wt	label claim	amt found	% labele caliam
pre1	0.221	0.222	0.145	0.145	0.145	5	4.977	99.54954955
pre 2	0.218	0.222	0.145	0.145	0.145	5	4.910	98.1981982
pre3	0.215	0.222	0.145	0.145	0.145	5	4.842	96.84684685
pre4	0.219	0.222	0.145	0.145	0.145	5	4.932	98.64864865
pre5	0.212	0.222	0.145	0.145	0.145	5	4.775	95.4954955
pre 6	0.217	0.222	0.145	0.145	0.145	5	4.887	97.74774775

Table 5: Results of Method precision for olopatadine by using uvmethod

Intermediate precision

The intermediate precision of the proposed method was determined by performing the method by two analysts (Analyst 1 and Analyst 2) for concentration of sample

solutions 100μ g/ml. The percent relative standard deviation (% RSD) for peak responses was calculated. The results for intermediate precision were shown in table 6.

	ANALYST 1		ANALYST 2	
S.NO	RT (MIN)	AREA (MV.SEC)	RT (MIN)	AREA(MV.SEC)
1	2.893	1389879	2.884	1385965
2	2.901	1397256	2.875	1377715
3	2.871	1372689	2.912	1366121
4	2.869	1377661	2.819	1345688
5	2.877	1388821	2.902	1366127
6	2.903	1401127	2.833	1399910
MEAN	2.885	1387905.5	2.8705	1373587.667
SD	0.014463748	10986.0312	0.037022966	17092.40841
%RSD	0.50134307	0.791554699	1.128774111	1.244362397
	ana	alyst 1 anal	yst 2	
Ċ	lay1 0	.229 0.2	221	
Ċ	lay2 0	.225 0.2	224	

Table 6: Results of Intermediate precision for olopatadine by using uv method

Acuracy

Accuracy of the method was confirmed by the standard addition method, which was carried out by performing recovery studies at 3 different concentrations 100%, 150% and 200% of these expected, in accordance with ICH guidelines, by replicate analysis (n=3). Known

amount of standard drug solution $(100\mu g/ml)$ was added to a pre analyzed sample solution $(100, 150, 200 \mu g/ml)$ and percentage drug content was measured. The closeness of obtained value to the true value indicates that the proposed method is accurate. Recovery studies were shown in table 7.

$$%$$
Recovery = [(Ct –Cpa)/Cs] × 100.

Where, Ct = Total concentration of analyte

Cpa = Concentration of pre-analysed sample

Cs = Concentration of standard added to pre-analysed sample.

Table 7: Results of recovery studies for olopatadine by using rp –hplc method						
s.no	level	std	Amount added	Total recovery	recovered	%recovery
1	50	100	50	150	50	100
2	50	100	50	150.057	50.057	100.114
3	50	100	50	150.78	50.78	101.56
4	100	150	100	200.421	100.421	100.421
5	100	150	100	200.602	100.036	100.326
6	100	150	100	247.117	147.117	98.078
7	100	150	150	247.271	147.271	98.18066
8	100	150	150	247.067	147.067	98.044

s.no	STND(µg/ml)	test (µg/ml)	absorbence at 217nm
1	25	20	0.444
2	25	25	0.485
3	25	30	0.512

Table 7: Results of recovery studies for olopatadine by using uvmethod

Robustness

The robustness study was performed to evaluate the influence of small but deliberate variation in the chromatographic condition. The robustness was checked by changing parameters like flow rate of mobile phase and detection wavelength

- Change in the detection wavelength by ± 2nm (294nm and 290nm)
- Change in flow rate by ± 0.1 ml/minute (1.6 ml/min and 1.4 ml/minute)

After each change, sample solution was injected and % assay with system suitability parameters were checked. Robustness values were given in table 8

Table 8: Results of Robustness for olopatadine by using rp - hplc method

parameter	Rt(min)	Area(mvsec)
Flow rate(ml/min)1.3	2.7	1485965
1.1	3.2	1300145
Wavelength(nm)252	2.78	1328815
256	2.76	1215141

WAVELENGTH	ABSORBENCE					
215	0.217	0.219	0.217	0.218	0.22	0.219
219	0.215	0.216	0.218	0.219	0.217	0.218

Limit of Detection and Quantitation

Detection and Quantitation limit were calculated by the method based on the standard deviation (σ) and slope of the calibration plot, using the formula Limit of Detection = $\sigma \times 3.3/S$ Limit of Quantitation = $\sigma \times 10/S$

Where σ = the standard deviation of the response. S = the slope of the calibration curve (of the analyte). Results of LOD & LOQ were shown in table 9.

Table 9: Results of LOD, LOQ for olopatadine		
S.No	LOD	LOQ
1	0.024982	0.075702

By using uv 2. 0.518571 1.571429

Specificity

Specificity of an analytical method is its ability to measure the analyte accurately and specifically in the presence of component that may be expected to be present in the sample matrix. Chromatograms of standard and sample solutions were compared in order to provide an indication of specificity of the method.

Assay of pharmaceutical formulation

The proposed validated method was successfully applied to determine olopatadine in their pharmaceutical dosage form And the % Assay results were shown in table 10.

Table 10: Results of % assay by using RP – HPLC method

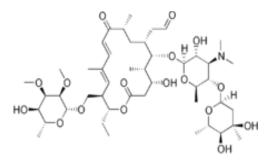
S.No	Amount Found	%Assay
1	5.0314	100.628
2	5.03590	100.718
3	5.0357	100.715

DRUG PROFILE

TYLOSIN TARTRATE

Drug substance: tylosin tartrate Category: anti bacterial agent.

Chemical structure



IUPAC name

 $[(2R,3R,4E,6E,9R,11R,12S,13S,14R) -12 - \{[3,6-dideoxy-4-O-(2,6-dideoxy-3-C-methyl-\alpha-L-ribo-hexopyranosyl) -3-(dimethylamino)-\beta-D-glucopyranosyl]oxy\}-2-ethyl-14-hydroxy-5, 9,13-trimethyl-8, 16-dioxo-11- (2-oxoethyl) oxacyclohexa deca-4,6-dien-3-yl] methyl 6-deoxy-2,3-di-O-methyl-\beta-D-allopyranoside.$

Molecular Formula: C 46H 77NO 17

Molecular Mass: 916.10g/mol

Synonyms: TYLOSIN TARTRATE SALT;TYLOSINE TARTRATE;pharmasin; Tylosin, [R-(R*,R*)] -2,3-dihydroxybutanedioate (salt);Tylosin, (2R,3R)-2,3-dihydroxybutanedioate (salt);TYLOSINTARTRATE,BP;

PHYSICAL PROPERTIES

Colour: clear, yellow-green State / form: powder form Solubility: freely soluble in water, dichloro methane.slightly soluble in alchol. Boiling Point(°C): 1116.3°C at 760 mmHg pka:7.3

MECHANISM OF ACTION

Tylosin tartate targets primarily gram positive bacteria and species of the Mycoplasma genus.categorised under macrolide antibiotics. Macrolide antibiotics inhibit bacterial growth by targeting the 50S ribosomal subunit preventing peptide bond formation and translocation during protein synthesis. Resistance to tylosin tartate is commonly attributed to mutations in 50S rRNA preventing tylosin tartate binding allowing the cell to synthesize proteins free of error. Tylosin is a macrolide, bacteriostatic antibiotic. It is similar in structure, mechanism of action, and spectrum as that of erythromycin. Tylosin is a macrolide antibiotic related structurally to erythromycin, tylosin is produced from Streptomyces fradiae. It is made naturally by the bacterium "Streptomyces fradiae" and acts to inhibit bacterial protein synthesis by inhibiting the 50S ribosome, a cellular structure only certain bacteria have and use to make internal proteins. It occurs as an almost white to buff-colored powder with a pKa of 7.1. It is slightly soluble in water and soluble in alcohol. Tylosin is considered to be highly lipid soluble. The tartrate salt is soluble in water. The injectable form of the drug (as the base) is in a 50% propylene glycol solution.

Tylosin is an antibiotic veterinary drug for the treatment of disease in food producing animals, including cattle, swine, and poultry, and for growth promotion in pigs. In small animals, tylosin is used for its anti-inflammatory properties in the large intestine rather than for its ability to fight infection. While few formal studies have been performed to examine this non-antibacterial property of tylosin, this has not stopped tylosin from being widely used in the treatment of colitis in pets. Tylosin is a broad spectrum antibiotic; it has good anti-bacterial activity against most pathogenic organism such as gram positive bacterium, some gram negative bacterium, vibrio, spirochete,

coccidian, etc. It is one of the first-choice drugs against infections caused by mycoplasma. Tylosin is fast acting; it can be absorbed quickly after oral administration and distributed widely in the organs (mainly concentrated in lung, liver, kidney, spleen and gallbladder). Bioavailability of Tylosin is high, which makes Tylosin's action rapid and long lasting. However, Tylosin is very safe. And residue in tissues is very rare. It is eliminated from the body upon Chemical Formula: C46H80N2O13

Molecular Weight: 869.15

Cas No.: 108050-54-0

withdrawal of the drug. Tylosin is very stable, against environmental conditions, feed processing, etc. Tylosin is a good growth promotant. It cannot only prevent diseases, but continual use also increases growth rate and can improve breeding efficiency. Because of Tylosin's broad spectrum activity it is quite effective when used as the only antibiotic for animal and poultry thus, cross-drug-resistance is avoided.

Specification: Tilmicosin consists of 82-88% cis isomer and 12-18% trans isomer, as determined by liquid chromatographic assay. (Assay: Min. 85%)

CONCLUSION

A simple, rapid, accurate, and precise RP-HPLC method for the analysis of olopatadine in pure and in pharmaceutical dosage forms had been developed and validated in accordance with ICH guidelines. The RP-HPLC method developed is cost-effective due to short retention time which enabled analysis of olopatadine samples with a small amount of mobile phase. From the % RSD values of precision and recovery studies the

method was found to be precise and accurate. The low detection and quantification limits achieved indicate the method is very sensitive. The robustness data gathered during method validation showed that the method is not susceptible to small changes in chromatographic conditions. The proposed RP-HPLC method developed by the author is suitable for routine analysis and quality assessment of olopatadine in pharmaceutical products.

Table 12: Summary of validated parameters for proposed method

Parameter	Result
Linearity range	$10-250 \ \mu g/ml$
Regression equation	Y =5844 x +75913
Slope	5844
Intercept	75913
Correlation coefficient	0.998
System precision (% RSD, n=5)	0.58197
Intermediate precision (% RSD, n=5)	1.2443
LOD (µg/ml)	0.099
LOQ (µg/ml)	0.075702
% Recovery (Accuracy, n =3)	98.044%
% Assay (% Assay, n=3)	100.365%

REFERENCES

- [1] Parsons CG, Danysz W, Quack G. Memantine is a clinically well Tolerated N-methyl-D-aspartate (NMDA) receptor antagonist-a review of preclinical data. Neuropharmacology. 38; 1999: 735-767.
- [2] Sonkusare SK, Kaul CL, Ramarao P. Dementia of Alzheimer's disease and other neurodegenerative disorders-memantine. Pharmacol Res. 51; 2005: 1-17.

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- [3] Erickson CA, Posey DJ, Stigler KA, Mullett J, Katschke AR, Mc Dougle CJ. A retrospective study of memantine in children and adolescents with pervasive developmental disorders. Psychopharmacology. 191; 2007: 141-147.
- [4] Zdanys K, Tampi RR. A systematic review of off-label uses of memantine for psychiatric disorders.Prog Neuropsychopharmacol Biol Psychiatry. 32; 2008: 1362-1374.
- [5] R F Suckow, M F Zhang, E D Collins, M W Fischman, T B Cooper. Sensitive and selective liquid chromatographic assay of memantine in plasma with fluorescence detection after pre-column derivatization. J Chromatogr B Biomed Sci Appl. 729 (1-2); 1999: 217-224
- [6] Afshin Zarghi, Alireza Shafaati, Seyed Mohsen Foroutan, Arash Khoddam, and Babak Madadian. Sensitive and Rapid HPLC Method for Determination of Memantine in Human Plasma Using OPA Derivatization and Fluorescence Detection. Application to Pharmacokinetic Studies. Sci Pharma. 78(4); 2010: 847–856.
- [7] Belen Puente, Esther Hernandez, Susana Perez, Luis Pablo, Esther Prieto, Maria Angeles Garcia, And Miguel Angel Bregante. Determination of memantine in plasma and vitreous humour by HPLC with precolumn derivatization and fluorescence detection. Journal of Chromatographic Science. 49; 2011: 745-752
- [8] Leis HJ, Fauler G, Windischhofer W. Quantitative analysis of memantine in human plasma by gas chromatography/negative-ion chemical ionization/mass spectrometry. J Mass Spectrom 37; 2002: 477-480.
- [9] K. Siddappa, Metre Mallikarjun, Tambe Mahesh, Kote Mallikarjun, Reddy Chandrakanth. Development and validation of a gas chromatographic method for the assay of memantine hydrochloride in pure and
